

Cyst Wall Biosynthesis Inhibitors: Discovery of a New Class of Anti-giardia Agents

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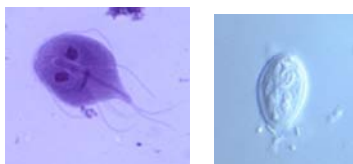
Introduction

Giardia lamblia

- Unicellular flagellated protozoal parasite
- Spread via contaminated food and water
- Giardia* is the most commonly diagnosed parasitic cause of diarrhea in North America
- Estimated 100 million cases of giardiasis worldwide
- Promiscuous: a wide variety of vertebrates can be infected
- Contact with companion animals and livestock is a risk factor for *giardiasis* in humans

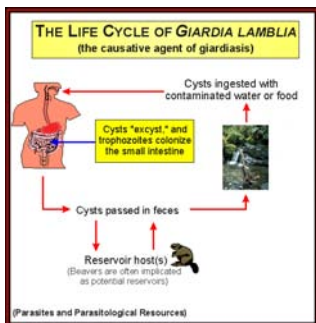
Giardia Lamblia Trophozoite

Giardia Lamblia Cyst



Treatment of Giardiasis

- Resistance to all anti-parasitics has been reported
- Failure rates of ~20% reported with currently used meds
- Cysts are highly resistant to chemical treatment
- >10% bleach or a mixture of 2% sulfuric acid and 2.5% potassium dichromate is needed to fully inactivate cysts
- Ozone, a new method to treat water supplies is not very effective
- Difficult to detect due to small size

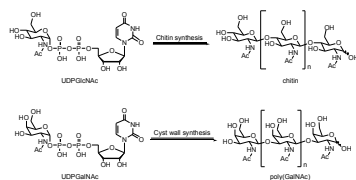


- Encystation is thought to be a response to unfavorable growth conditions and allows survival outside the host
- Encystation is common to many protozoans

Cyst Wall Synthesis: a New Target for Anti-Protozoal Therapy

- Many protozoal cyst walls are composed of chitin or a chitin like substance
 - Giardia*: β-1-3-linked poly(N-acetylglucosamine) [poly(GalNAc)]
 - Entamoeba*: chitin
- Enzyme activity in *Giardia* termed cyst wall synthase has been described¹
- An inhibitor of cyst wall synthesis is likely to be a broad spectrum anti-protozoal agent

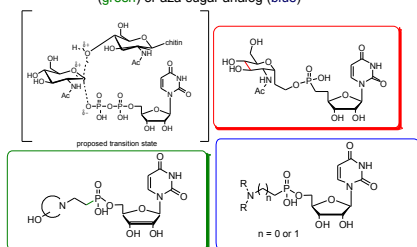
Chitin and cyst wall synthesis



Inhibitor Design

We have designed and synthesized two classes of inhibitors of protozoal cyst wall synthesis

- A phosphonate C-glycoside UDP-glucosamine analog that is metabolically, chemically stable and is able to penetrate cell membranes (below, red)
- Transition-state analogs containing an aza-sugar (green) or aza-sugar analog (blue)



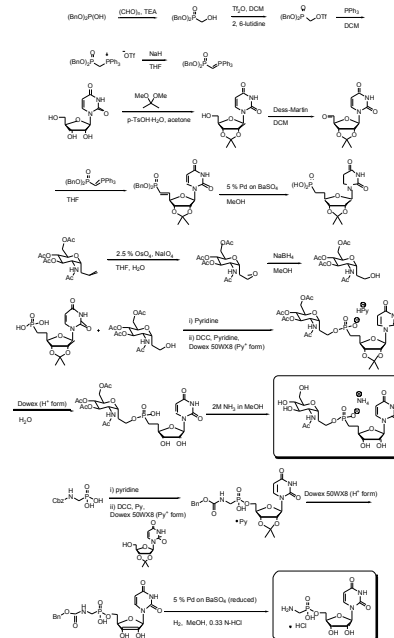
Biological Assays

- Anti-Giardia**
 - The *G. lamblia* WB-C6 strain was used for the assays.
 - Trophozoite inhibition was determined by culturing the parasites anaerobically for 48 hours in the presence of 2-fold drug dilutions in 96-well plates in duplicate. The XTT reagent was used to measure cell viability.
 - Cyst development was determined in a similar manner except the assay was conducted for 4 days in encysting medium in glass vials followed by counting cysts, washing cells in dH₂O and resuspension in trophozoite culture medium to evaluate cyst viability.
- Host toxicity**
 - Manine Darbey Bovine Kidney cells (MDBK) were used to evaluate mammalian cell toxicity. 2-fold drug dilutions in RPMI1640 culture medium with 5% FBS were used with cells seeded and grown for 48 hours before performing viability measurements using the XTT reagent.
- IC₅₀ determination**
 - Standard curves were generated as % no drug controls for each drug dilution. At least 3 independent determinations were used to determine the values.

Anti-Giardia Activity

Structure	Giardia MIC (μM)	MDBK Cell Inc 50% (μM)	Therapeutic index	Giardia cyst formation percent of control
	3.9	>250	64	27.5
	15.6	60	10.7	9.7
	3.9	>125	32	56.2
	>20	>250		26.4
	1.9	>250	131	10.3
	>20	>250		34.4
	>10	>250		14.9
	>10	>250		72.2
	>10	>250		36.6
	>10	>250		27.5
	0.48	>250	>520	5.73
	>20	>250		28.7
	>20	>250		22.0
	>20	>250		17.2
	5			13.7

Synthesis of Selected Targets



Conclusion

We have designed and synthesized a new class potent anti-Giardia agents that:

- Have very low toxicity
- Have micromolar to sub-micromolar MIC against *Giardia* trophozoite growth
- We have discovered an agent that is a highly promising lead for clinical development.
- Ten times more potent than metronidazole (MIC 0.48 μM vs 5μM)
- Has very low toxicity
- Enzyme assays are in progress
- Assays against other protozoans are in progress
- Animal model studies will begin in due course

References

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